21. Nucleotides

Part XLI1)

The 2-Dansylethoxycarbonyl

(= 2-{[5-(Dimethylamino)naphthalen-1-yl]sulfonyl}ethoxycarbonyl; Dnseoc) Group for Protection of the 5'-Hydroxy Function in Oligodeoxyribonucleotide Synthesis

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Use of the 2-dansylethoxycarbonyl (= 2-{[5-(dimethylamino)naphthalen-1-yl]sulfonyl}ethoxycarbonyl; Dnseoc) group as an intermediate 5'-OH protecting group in oligodeoxyribonucleotide synthesis using the automated phosphoramidite approach is described in a model study to an alternative strategy in RNA synthesis.

1. Introduction. – The development of an adequate protecting-group combination especially for the 2'- and 5'-OH functions is still a substantial problem in oligoribonucle-otide synthesis. The use of acid-labile 2'-OH protecting groups, e.g. the tetrahydro-2H-pyranyl [2-4] and tetrahydro-4-methoxy-2H-pyran-4-yl groups [5-7], requires the displacement of the traditional trityl blocking groups [8-10] at the 5'-OH function due to unsatisfactory compatibility [11-13], despite the fact that recently some progress was achieved by applying the 1-(2-chloro-4-methylphenyl)- [14-16] and 1-(2-fluoro-phenyl)-4-methoxypiperidin-4-yl (Ctmp and Fpmp, resp.) protecting groups [17-19] in combination with the pixyl group. As base-labile functions, the levulinyl (Lev) group [20-24] and (fluoren-9-yl)methoxycarbonyl (Fmoc) group [25-27] were mainly suggested. The Fmoc group was also used in oligodeoxynucleotide synthesis to guarantee non-depurinating conditions [28-31].

To improve this alternative strategy, we have developed the 2-dansylethoxycarbonyl (= 2-{[5-(dimethylamino)naphthalen-1-yl]sulfonyl}ethoxycarbonyl; Dnseoc) group as a new base-labile 5'-OH protecting group in oligonucleotide synthesis. The Dnseoc group can easily be cleaved with dilute 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in aprotic solvents by β -elimination. The condensation yield in solid-phase syntheses can be determined by UV detection at 350 nm or, more sensitively, by fluorescence detection at 530 nm of the resulting 5-(dimethylamino)naphthalen-1-yl vinyl sulfone.

¹⁾ Part XL: [1].

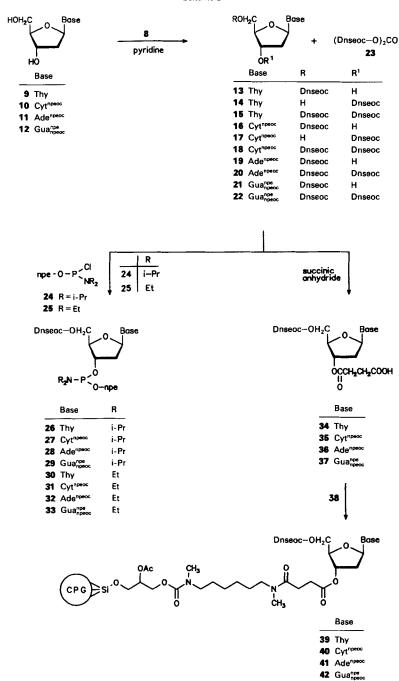
In this paper, we wish to report the usefulness of the Dnseoc protecting group in oligodeoxyribonucleotide synthesis as a model study on the way to a new approach in oligoribonucleotide synthesis.

2. Syntheses. – To introduce the Dnseoc group into the 5'-O-position of a nucleoside, 2-dansylethyl chloroformate hydrochloride (8) was used. This reagent 8 was synthesized starting from 5-aminonaphthalene-1-sulfonic acid (1) by known but partially improved procedures [32–34] (Scheme 1). The amino function of 1 was first methylated with dimethylsulfate and the resulting 5-(dimethylamino)naphthalene-1-sulfonic acid (2) chlorinated with PCl₅ to yield dansyl chloride (3) according to Horner and Lindel [32]. After reduction of 3 with Na₂SO₃ to 5-(dimethylamino)naphthalene-1-sulfinic acid (4) and transformation into its corresponding sodium salt 5, reaction with 2-chloroethanol gave 2-dansylethanol (6) according to a procedure of Goya et al. [33]. As a side product, 1,2-bis(dansyl)ethane (7) could be isolated in 9% yield which was not yet described and may have been formed from 5-(dimethylamino)naphthalene-1-yl vinyl sulfone and 5. Treatment of 6 with trichloromethyl chloroformate in MeCN led to 8 of correct analytical composition, whereas the procedure of Goya et al. [34] turned out to be not reproducible and did not give pure material, despite the fact that these authors claim a correct elemental analysis.

As protecting groups for the functional residues of the nucleobases, the already established 2-(4-nitrophenyl)ethoxycarbonyl (npeoc) and 2-(4-nitrophenyl)ethyl (npe) groups were used [35] [36]. Best results to introduce the Dnseoc group were obtained by adding a slight excess of solid 8 at 0° to a pyridine solution of the 2'-deoxynucleosides 9–12. The reaction was complete within 1 h, and after workup and flash chromatography, the desired 5'-O-Dnseoc-nucleosides 13, 16, 19, and 21, respectively, were obtained in 65–80% yield, besides small amounts of 3',5'-di-O-substituted nucleosides 15, 18, 20, and 22, respectively (8–15%). The 3'-O-Dnseoc-nucleosides could be isolated only in case of the thymidine and cytidine derivatives (5% of 14 and 8% of 17, resp.). A by-product in this reaction was always bis(2-dansylethyl)carbonate (23) which could be formed in yields up to 20%.

The 3'-phosphoramidites 26–33 were synthesized by phosphitylation using 2-(4-nitrophenyl)ethyl chloro(diisopropylamido)phosphite (24) and 2-(4-nitrophenyl)ethyl chloro-(diethylamido)phosphite (25) [37] [38]. The yields after workup and flash chromatography (silica gel) ranged from 45 to 68%.

Scheme 2



Solid-phase synthesis of oligodeoxyribonucleotides via the npe/npeoc approach [38–40] requires a DBU-stable linkage of the starting nucleoside through a spacer molecule to a glass-bead support. Therefore, the 5'-O-Dnseoc-3'-O-succinyl-nucleosides 34–37 were synthesized by reaction of 13, 16, 19, and 21 with succinic anhydride and 4-(dimethylamino)pyridine in CH₂Cl₂ [41] in almost quantitative yields. The 3'-O-succinyl-nucleosides 34–37 were then reacted with LCAMA-CPG (= (long-chain-alkyl)methylamine controlled-pore glass; 38) [38–40] using the coupling reagent O-{[cyano(ethoxycarbonyl)methylidene]amino}-1,1,3,3-tetramethyluronium tetrafluoroborate (TOTU) [42] and N-methylmorpholine in MeCN followed by a capping process with Ac₂O and 4-(dimethylamino)pyridine in pyridine (\rightarrow 39–42). Loadings of ca. 20 μ mol/g were reached with a 500 Å-CPG material.

The assembly of oligodeoxyribonucleotides was carried out by the solid-phase phosphoramidite method similar to general procedures [43-46]. Four chemical steps and intermediate washing steps were performed using an automated DNA synthesizer by sequential passage of reagents through a column containing the nucleoside-functionalized support (39-42): 1) deprotection of terminal Dnseoc groups with 0.1M DBU in MeCN for 140 s; 2) coupling with 0.1m nucleoside phosphoramidite (26-33) and 0.5m 1H-tetrazole in MeCN for 40-60 s; 3) capping with Ac₂O/2,6-dimethylpyridine/1methyl-1H-imidazole in THF for 50 s; 4) oxidation with 0.05M I, in THF/pyridine/ H,O for 53 s. The completeness of each coupling step was checked by measurement of the absorbance at 350 nm of the eluate from the Dnseoc-deprotection step 1. The determination of condensation yields was also possible by measuring the fluorescence at 530 nm. After the last synthesis cycle, the support was treated with 1 m DBU in MeCN for 10 h to remove all npeoc and npe protecting groups. Finally, the fully deblocked oligodeoxyribonucleotide was cleaved from the support by treatment with concentrated NH₃ solution for 200 min. Thereafter, the oligonucleotide-containing ammoniacal solution from which the amount of isolated oligonucleotide was determined by measurement of the absorbance at 260 nm was lyophilized in a Speed-vac concentrator. The quality of the synthesized oligonucleotides was proven by reversed-phase and anion-exchange HPLC. Table 1 shows the synthesized oligodeoxyribonucleotides and the isolated yields.

Sequence	Amount of starting nucleoside [µmol]	Isolated yield [OD ₂₆₀]	
(dT) ₁₀	0.6	39.2	
(5'-3')[d(A-A-A-T)]	0.45	18.0	
(5'-3')[d(A-A-A-A-A-T)]	0.56	25.0	
$(dA)_{10}$	0.64	48.0	
(5'-3')[d(C-C-C-C-T)]	0.5	17.0	
$(dC)_{10}$	0.6	22.4	
(5'-3')[d(G-G-G-G-G-G-T)]	0.6	28.0	

Table 1. Synthesized Oligodeoxyribonucleotides

The N,N-diethylphosphoramidites turned out to be more reactive than the corresponding N,N-diisopropylphosphoramidites, and, therefore, more careful handling due to a higher sensitivity to moisture was required. The efficiency of this new method is demonstrated by the anion-exchange HPLC of 'crude' (dT)₁₀ isolated without any further

Table 2. Physical Data of Nucleoside Derivatives

	UV Spectra	(MeOH)							
	λ _{max} [nm]			lg ε					
13	214, 257, 346			4.66, 4.32, 3	4.66, 4.32, 3.60				
14	215, 258, 34	215, 258, 346			4.64, 4.36, 3.61				
15	216, 255, 34	6		4.87, 4.55, 3.91					
16	212, 246, 28	212, 246, 286 (sh), 343			4.81, 4.48, 4.19, 3.63				
17	213, 246, 286 (sh), 343			4.83, 4.50, 4.21, 3.65					
18	213, 249, 286 (sh), 346			5.06, 4.68, 4.25, 3.94					
19	211, 263, 343			4.83, 4.54, 3	4.83, 4.54, 3.61				
20	213, 259, 343			5.03, 4.70, 3	5.03, 4.70, 3.94				
21	214, 262, 341			4.91, 4.63, 3	4.91, 4.63, 3.67				
22	214, 256, 34	3		5.09, 4.76, 3	3.90				
26	205, 260, 34								
27	212, 254, 28	5 (sh), 342		4.87, 4.54, 4	1.37, 3.61				
28	206, 265, 341			4.95, 4.67, 3	3.65				
29	213, 264, 33								
30	213, 260, 34	3		4.74, 4.47, 3	4.74, 4.47, 3.61				
31	212, 255, 28-	55, 284 (sh), 340 4.85, 4.54, 4.38, 3.64							
32	210, 264, 34	3		4.89, 4.66, 3.65					
33	214, 264, 34	0		4.93, 4.70, 3.66					
34	214, 257, 34	214, 257, 346			4.71, 4.36, 3.63				
35	212, 247, 286 (sh), 343			4.79, 4.47, 4.18, 3.61					
36	211, 263, 342			4.83, 4.57, 3.64					
37	214, 262, 34	0		4.89, 4.62, 3	3.66				
	¹ H-NMR Spectra (CDCl ₃ , δ [ppm])								
	H-C(1')	H-C(2')	H'-C(2')	H-C(3')	H-C(4')	H-C(5')	H-C(5")		
13	6.29(t)	2.38(m)	2.13(m)	4.32(m)	4.02(m)	4.2	20 (m)		
14	6.08 (dd)	2.46(m)	2.30(dd)	5.14(m)	3.97(m)	3.8	85 (m)		
15	6.24(dd)	2.34(m)	2.07(m)	4.96(d)	3.98(m)	4.31 (dd)	4.22(dd)		
16	6.24(t)	2.66(m)	2.08(m)	a)	4.14(m)	[4.30 to	4.19]* (m		
17	6.04(t)	2.49	(m)	5.14(m)	4.06(m)	3.91(m)	3.81(m)		
18	6.15 (dd)	2.66 (dd)	1.98(m)	4.93(d)	4.08(m)	[4.35 to	4.22](m)		
19	6.47(t)	2.82(m)	2.56(m)	4.61(m)	4.16(m)	4.2	22 (m)		
20	6.40 (dd)	2.86(m)	2.55 (dd)	5.16(d)	4.15(m)	4.3	29 (m)		
21	6.43(t)	2.84(m)	2.51(m)	4.77(t)	4.16(m)	4.32(dd)	4.23 (dd)		
22	6.23(t)	3.05(m)	2.44(m)	5.27(m)	4.14(m)	[4.40 to	4.29](m)		
26 ^b)	6.12(t)	2.17(m)	1.92(m)	4.27(m)	[4.12	to	3.92](m)		
27 ^b)	6.05(t)	2.44(m)	1.95(m)	4.25(m)	[4.15	to	3.97(m)		
28 ^b)	6.32(m)	2.71(m)	2.46(m)	4.59(m)	[4.20	to	4.00](m)		
29 ^b)	6.19(m)	2.81(m)	2.36(m)	4.70(m)	[4.26	to	3.99] (m)		
30 ^b)	6.10(t)	2.17(m)	1.93(m)	4.28(m)	[4.12	to	3.95(m)		
31 ^b)	6.03 (m)	2.41 (m)	1.95(m)	4.25 (m)	[4.15	to	3.97(m)		
31)	6.30(t)	2.71 (m)	2.45(m)	4.59(m)	[4.18	to	4.00](m)		
	(-)	2.79 (m)	2.35 (m)	4.75 (m)	[4.25	to	3.97] (m)		
32 ^b)	6.17(m)		(' /	, .	-	4.15(m)			
32 ^b) 33 ^b)	6.17(m) $6.14(t)$		S(t)	5.09(m)	4.02(m)	4	15 (m)		
32 ^b) 33 ^b) 34 ^c)	6.14(t)	2.23	* /	5.09(m) 5.10(m)	4.02 (m) 4.16 (m)		` '		
32 ^b) 33 ^b) 34 ^c) 35 ^c)	, ,		2.17 (m) 2.56 (m)	5.09 (m) 5.10 (m) 5.35 (m)	4.02 (m) 4.16 (m) [4.24		15 (m) 16 (m) 4.13] (m)		

^a) Chemical shift is in area marked with *. ^b) In CD₃CN. ^c) In (D₆)DMSO.

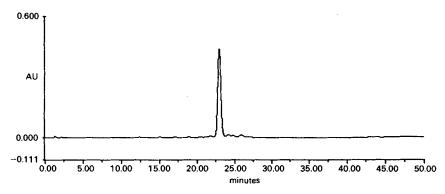


Figure. Anion-exchange HPLC of $(dT)_{10}$. Column: Nucleogen 60-7 DEAE, 7 µm, 4×125 mm (Macherey-Nagel); gradient: 0-0.6M LiCl (0-50 min) in 20% MeCN, 0.02M NaOAc pH 7.5; flow: 1 ml/min.

purification process (Fig.). The positive results can also be seen as an obvious encouragement to extend the strategy to RNA synthesis.

3. Physical Data. – The structural assignments of the newly synthesized nucleoside derivatives are based on elemental analyses and UV and 'H-NMR spectra. The UV spectra show a new bathochromic absorption at 340 nm due to the dansyl chromophore. The 'H-NMR spectra are of complex nature due to the variety of blocking groups. The characteristic sugar protons are listed in *Table 2*. A shift to lower field indicates the position of substitution.

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Experimental Part

General. Products were dried under high vacuum. TLC: Precoated silica gel thin-layer sheets F1500 LS 254 from Schleicher & Schuell. Flash chromatography (FC): silica gel (Baker, 30–60 µm); 0.2–0.3 bar. M.p.: Gallenkamp melting-point apparatus; no corrections. UV/VIS: Perkin-Elmer, Lambda 15; λ_{max} in nm (log ε). ¹H-NMR: Bruker AC 250; in ppm rel. to TMS or CDCl₃ ((D₆)DMSO, CD₃CN) as internal standard. ³¹P-NMR: Joel 400 MHz; in ppm rel. to H₃PO₄.

- 1. 5-(Dimethylamino) naphthalene-1-sulfonic Acid (2) [32]. In a 3-1 Erlenmeyer flask, 150 g (0.66 mol) of 5-aminonaphthalene-1-sulfonic acid (1) were added slowly in portions to 210 g (2.5 mol) of NaHCO₃ in 450 ml of H₂O (strong CO₂ evolution). Then, 150 ml (1.52 mol) of dimethyl sulfate were added dropwise within 30 min to the stirred ice-cooled soln. which was warmed to 80° for 30 min and after cooling acidified with 75 ml of conc. HCl soln. (pH 4). The precipitated product was filtered, washed with a small quantity of H₂O and dried to constant weight in the air and then at 120°: 148.7 g (90%) of gray powder. M.p. $> 300^{\circ}$.
- 2.5-(Dimethylamino) naphthalene-1-sulfonyl Chloride (= Dansyl Chloride; 3) [32]. Under ice-cooling (exothermic reaction), 80 g (0.32 mol) of 2 and 70 g (0.34 mol) of PCl₅ were mixed with a glass rod. To complete the reaction, the mixture was warmed to 60° for 2 h under exclusion of moisture. Then it was poured on 1 lice/H₂O. After careful neutralization with 140 g (1.67 mol) of NaHCO₃, the product was extracted with Et₂O (5 × 500 ml), the org. layer dried (Na₂SO₄) and evaporated, and the residue purified by FC (silica gel, 20×5.5 cm, 600 ml AcOEt): 64.9 g (75%) of 3. Orange-yellow crystals. M.p. 69° ([32]: m.p. 69°). 1 H-NMR (CDCl₃): 8.70 (d, H-C(2)); 8.43, 8.36 (2d, H-C(4), H-C(8)); 7.70, 7.58 (2t, H-C(3), H-C(7)); 7.27 (d, H-C(6)); 2.91 (s, Me₂N).

- 3. 5-(Dimethylamino) naphthalene-I-sulfinic Acid (4) [33]. A suspension of 120 g (0.95 mol) of Na₂SO₃ in 200 ml of H₂O was weakly alkalinized with 1 ml of 40% NaOH, then 40 g (0.15 mol) of 3 were added. The mixture was stirred at r.t. for 2.5 h. After dilution with H₂O (200 ml) and filtration, the product was precipitated by slow addition of 28 ml of conc. H₂SO₄ soln. (pH 4). The beige crystals were recrystallized from H₂O: 31.8 g (85%) of bluish plates. M.p. 160–161° ([33]: m.p. 160°). Anal. calc. for $C_{12}H_{13}NO_2S \cdot 1 H_2O$ (253.3): C 56.90, H 5.97, N 5.53; found: C 56.52, H 5.91, N 5.51.
- 4. 2-Dansylethanol (= $2-\{[5-(Dimethylamino)naphthalen-1-yl]sulfonyl\}ethanol$; 6) [33] and 1,2-Bis(dansyl)ethane (= 1,2-Bis{[5-(dimethylamino)naphthalen-1-yl]sulfonyl}ethane; 7). To a soln. of 2.4 g (60 mmol) of NaOH in 45 ml of H₂O, 15 g (59.2 mmol) of 4 were added. Dissolution was completed with a few drops of 40% NaOH soln. Then, 15 ml of EtOH and 24 ml (0.36 mol) of 2-chloroethanol were added. The soln. was stirred at 80° for 20 h. The yellow precipitate was isolated by filtration from the hot mixture. Washing with a small quantity of MeOH and drying under high vacuum gave 1.02 g (7%) of 7. The cooled filtrate was extracted with CH₂Cl₂ (1 × 50 ml, 2 × 25 ml), the org. layer washed with H₂O (100 ml), the aq. phase extracted with CH₂Cl₂ (50 ml), the combined org. phase dried (Na₂SO₄) and evaporated, and the resulting green oil crystallized after addition of H₂O (50 ml). The yellow-green crystals were dried in a desiccator over P₄O₁₀ to give 10.71 g (65%) of 6 which was used in this form for further reactions. Recrystallization from H₂O yielded yellow needles.
- **6**: M.p. 100° ([33]: m.p. 100–101°). $pK_a = 3.42$. UV (MeOH): 215 (4.66), 252 (4.18), 342 (3.67). ¹H-NMR (CDCl₃): 8.63 (d, H-C(2)); 8.33 (2d, H-C(4), H-C(8)); 7.61 (2t, H-C(3), H-C(7)); 7.22 (d, H-C(6)); 3.97 (m, CH₂O); 3.56 (t, SO₂CH₂); 2.90 (t, Me₂N); 2.88 (br., OH). Anal. calc. for C₁₄H₁₇NO₃S (279.4): C 60.19, H 6.13, N 5.01; found: C 60.10, H 6.22, N 5.14.
- 7: M.p. 218–220°. UV (CHCl₃): 257 (4.45), 354 (3.91). 1 H-NMR (CDCl₃): 8.61 (*d*, 2 H, H–C(2)); 8.20 (*t*, 4 H, H–C(4), H–C(8)); 7.61–7.53 (*m*, 4 H, H–C(3), H–C(7)); 7.22 (*d*, 2 H, H–C(6)); 3.66 (*s*, CH₂CH₂); 2.91 (*s*, 2 Me₂N). Anal. calc. for $C_{26}H_{28}N_{2}O_{4}S_{2}$ (496.7): C 62.88, H 5.68, N 5.64; found: C 62.36, H 5.77, N 5.57.
- 5. 2-Dansylethyl Chloroformate Hydrochloride (= 2-{[5-(Dimethylamino)naphthalen-1-yl]sulfonyl}ethyl Chloroformate Hydrochloride; 8). Under ice-cooling and stirring, 2.4 ml (19.9 mmol) of trichloromethyl chloroformate were added to 30 ml of dry MeCN. Then, a soln. of 3 g (10.7 mmol) of 6 in 15 ml of dry MeCN was added dropwise into the stirred and ice-cooled mixture within 10 min and stirring continued for another 5 h. The colourless precipitate was collected, washed with dry THF, and dried under high vacuum: 3.70 g (91%) of 8. M.p. 154–155°. UV (MeOH): 215 (4.62), 252 (4.14), 345 (3.62). ¹H-NMR ((D₆)DMSO): 9.4 (br., NH); 9.06–7.84 (d, t, 3m, 6 arom. H); 3.66 (m, CH₂CH₂); 3.21 (d, Me₂N). Anal. calc. for C₁₅H₁₆ClNO₄S·HCl (378.3): C 47.63, H. 4.53, N 3.70; found: C 47.90, H 4.64, N 3.96.
- 6. 5'-O-(2-Dansylethoxycarbonyl) thymidine (13), 3'-O-(2-Dansylethoxycarbonyl) thymidine (14), and 3',5'-Bis-O-(2-dansylethoxycarbonyl) thymidine (15). In dry pyridine (2 × 30 ml), 2 g (8.26 mmol) of thymidine (9) were co-evaporated and then taken up in dry pyridine (80 ml). After cooling in an ice-bath, 8 (4.06 g, 10.73 mmol) was added in solid form. The ice-cooled mixture was stirred for 1 h, then the reaction stopped with MeOH (10 ml). After evaporation, the residue was diluted with CH_2Cl_2 (100 ml) and washed with sat. $NaHCO_3$ soln. (100 ml), the aq. phase extracted with CH_2Cl_2 (2 × 50 ml), the combined org. layer dried (Na_2SO_4), evaporated, and co-evaporated with toluene (2 × 50 ml), AcOEt/EtOH 1:1 (50 ml), and CH_2Cl_2 (30 ml), and the residue purified by FC (silica gel, 25 × 4 cm, $CH_2Cl_2/MeOH$ 100:0 (0.51), 99:1 (11), 98:2 (21; 15, then 14), 97:3 (0.51; 14), and 96:4 (0.81; 13)): 3.62 g (80%) of 13, 206 mg (5%) of 14, and 0.67 g (10%) of 15 as yellow amorphous solids.
- 13: UV (MeOH): 214 (4.66), 257 (4.32), 346 (3.60). 1 H-NMR (CDCl₃): 9.19 (br., NH); 8.60 (d, H–C(2)(Dns)); 8.33–8.28 (m, H–C(4)(Dns), H–C(8)(Dns)); 7.65–7.57 (m, H–C(3)(Dns), H–C(7)(Dns)); 7.22 (d, H–C(6)(Dns)); 7.21 (s, H–C(6)); 6.29 (t, H–C(1')); 4.64–4.39 (2m, CH₂O); 4.32 (m, H–C(3')); 4.20 (m, 2 H–C(5')); 4.02 (m, H–C(4')); 3.72 (m, SO₂CH₂); 3.60 (br., OH–C(3')); 2.89 (s, Me₂N); 2.42–2.33 (m, H–C(2')); 2.18–2.08 (m, H'–C(2')); 1.84 (d, Me). Anal. calc. for C₂₅H₂₉N₃O₉S·1 H₂O (565.6): C 53.09, H 5.52, N 7.42; found: C 52.61, H 5.23, N 7.24.
- 14: UV (MeOH): 215 (4.64), 258 (4.36), 346 (3.61). 1 H-NMR (CDCl₃): 8.90 (br., NH); 8.64 (d, H-C(2)(Dns)); 8.31 (2d, H-C(4)(Dns), H-C(8)(Dns)); 7.62 (2t, H-C(3)(Dns), H-C(7)(Dns)); 7.43 (s, H-C(6)); 7.22 (d, H-C(6)(Dns)); 6.08 (dd, H-C(1')); 5.14 (m, H-C(3')); 4.50 (t, CH₂O); 3.97 (m, H-C(4')); 3.85 (m, 2 H-C(5')); 3.72 (t, SO₂CH₂); 2.90 (s, OH-C(5'), Me₂N); 2.51-2.40 (m, H-C(2')); 2.34-2.25 (dd, H'-C(2')); 1.91 (s, Me). Anal. calc. for C₂₅H₂₉N₃O₉S (547.6): C 54.84, H 5.34, N 7.67; found: C 54.41, H 5.40, N 7.57.
- **15**: UV (MeOH): 216 (4.87), 255 (4.55), 346 (3.91). ¹H-NMR (CDCl₃): 8.82 (br., NH); 8.63 (2*d*, 2 H, H–C(2)(Dns)); 8.33–8.28 (*m*, 4 H, H–C(4)(Dns), H–C(8)(Dns)); 7.65–7.56 (*m*, 4 H, H–C(3)(Dns), H–C(7)(Dns)); 7.20 (*d*, 2 H, H–C(6)(Dns)); 7.19 (*s*, H–C(6)); 6.24 (*dd*, H–C(1')); 4.96 (*d*, H–C(3')); 4.62–4.42 (*m*, 2 CH₂O); 4.33–4.20 (2*dd*, 2 H–C(5')); 3.98 (*m*, H–C(4')); 3.71 (*m*, 2 SO₂CH₂); 2.89 (*s*, 2 Me₂N); 2.37–2.31 (*m*,

H–C(2')); 2.13–2.01 (m, H'–C(2')); 1.84 (s, Me). Anal. calc. for $C_{40}H_{44}N_4O_{13}S_2 \cdot 1$ H_2O (871.0); C 55.16, H 5.32, N 6.43; found: C 55.11, H 5.19, N 6.39.

7. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]cytidine (16), 3'-O-(2-Dansylethoxycarbonyl-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]cytidine (17), and 3',5'-Bis-O-(2-dansylethoxycarbonyl)-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]cytidine (18). As described in Exper. 6, with 2 g (4.76 mmol) of 10 [35] [36] and 2.34 g (6.18 mmol) of 8 in 50 ml of dry pyridine. Purification was achieved by FC (silica gel, 18×4 cm, $CH_2Cl_2/MeOH$ 100:0 (0.31), 99:1 (11), 98:2 (21); 18, then 17), 97:3 (0.51; 17), and 96:4 (0.51; 16)) to give 2.26 g (65%) of 16, 279 mg (8%) of 17, and 0.73 g (15%) of 18 as yellow amorphous solids.

16: UV (MeOH): 212 (4.81), 246 (4.48), 286 (sh, 4.19), 343 (3.63). 1 H-NMR (CDCl₃): 8.59 (*d*, H–C(2)(Dns)); 8.33–8.28 (*m*, H–C(4)(Dns), H–C(8)(Dns)); 8.15 (*d*, 2 H o to NO₂); 8.15 (br., NH); 7.92 (*d*, H–C(6)); 7.59 (2*t*, H–C(3)(Dns), H–C(7)(Dns)); 7.40 (*d*, 2 H m to NO₂); 7.20 (*d*, H–C(5), H–C(6)(Dns)); 6.24 (*t*, H–C(1')); 4.59–4.40 (*m*, 2 CH₂O); 4.30–4.19 (*m*, H–C(3'), 2 H–C(5'), OH–C(3')); 4.14 (*m*, H–C(4')); 3.72 (*t*, SO₂CH₂); 3.10 (*t*, C–CH₂); 2.88 (*s*, Me₂N); 2.72–2.60 (*m*, H–C(2')); 2.13–2.02 (*m*, H′–C(2')). Anal. calc. for C₃₃H₃₅N₅O₁₂S·1 H₂O (743.8): C 53.29, H 5.01, N 9.42; found: C 53.20, H 4.79, N 9.19.

17: UV (MeOH): 213 (4.83), 246 (4.50), 286 (sh, 4.21), 343 (3.65). 1 H-NMR (CDCl₃): 8.63 (d, H–C(2)(Dns)); 8.31 (2d, H–C(4)(Dns), H–C(8)(Dns)); 8.18 (d, 2 H o to NO₂); 8.08 (d, H–C(6)); 7.72 (br., NH); 7.66–7.58 (m, H–C(3)(Dns), H–C(7)(Dns)); 7.40 (d, 2 H m to NO₂); 7.22 (d, H–C(6)(Dns)); 7.17 (d, H–C(5)); 6.04 (t, H–C(1')); 5.14 (m, H–C(3')); 4.53–4.42 (m, 2 CH₂O); 4.06 (m, H–C(4')); 3.93–3.78 (m, 2 H–C(5')); 3.71 (t, SO₂CH₂); 3.11 (t, C–CH₂); 3.00 (br., OH–C(5')); 2.90 (t, Me₂N); 2.60–2.37 (t, 2 H–C(2')). Anal. calc. for C₃₃H₃₅N₃O₁₂S·1 H₂O (743.8): C 53.29, H 5.01, N 9.42; found: C 52.95, H 4.77, N 9.05.

18: UV (MeOH): 213 (5.06), 249 (4.68), 286 (sh, 4.25), 344 (3.94). 1 H-NMR (CDCl₃): 8.62 (2*d*, 2 H, H–C(2)(Dns)); 8.33–8.28 (*m*, 4 H, H–C(4)(Dns), H–C(8)(Dns)); 8.16 (*d*, 2 H o to NO₂); 7.85 (*d*, H–C(6)); 7.65–7.55 (*m*, 5 H, NH, H–C(3)(Dns), H–C(7)(Dns)); 7.37 (*d*, 2 H o to NO₂); 7.22–7.18 (*m*, 3 H, H–C(5), H–C(6)(Dns)); 7.21 (*s*, H–C(6)); 6.15 (*dd*, H–C(1')); 4.93 (*d*, H–C(3')); 4.54–4.41 (*m*, 3 CH₂O); 4.28 (*m*, 2 H–C(5')); 4.08 (*m*, H–C(4')); 3.69 (*m*, 2 SO₂CH₂); 3.10 (*t*, C–CH₂); 2.89, 2.88 (2*s*, 2 Me₂N); 2.70–2.61 (*dd*, H–C(2')); 2.04–1.92 (*m*, H′–C(2')). Anal. calc. for C₄₈H₅₀N₆O₁₆S₂·0.5 H₂O (1040.1): C 55.43, H 4.94, N 8.08; found: C 55.28, H 4.90, N 7.90.

- 8. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy- N^6 -[2-(4-nitrophenyl)ethoxycarbonyl]adenosine (19) and 3',5'-Bis-O-(2-dansylethoxycarbonyl)-2'-deoxy- N^6 -[2-(4-nitrophenyl)ethoxycarbonyl]adenosine (20). As described in Exper. 6, with 2 g (4.5 mmol) of 11 [35] [36] and 2.21 g (5.85 mmol) of 8 in 40 ml of dry pyridine. Purification was achieved by FC (silica gel, 18×4 cm, $CH_2Cl_2/MeOH$ 100:0 (0.3 l), 99:1 (11; 20), 98:2 (0.5 l; 20), 97:3 (1 l; 19)) to give 2.32 g (69%) of 19 and 0.47 g (10%) of 20 as yellow amorphous solids.
- **19**: UV (MeOH): 211 (4.83), 263 (4.54), 343 (3.61). 1 H-NMR (CDCl₃): 8.80 (br., s, NH); 8.67 (s, H–C(2)); 8.59 (d, H–C(2)(Dns)); 8.32–8.26 (m, H–C(4)(Dns), H–C(8)(Dns)); 8.13 (s, H–C(8)); 8.13 (d, 2 H o to NO₂); 7.63–7.55 (m, H–C(3)(Dns), H–C(7)(Dns)); 7.41 (d, 2 H m to NO₂); 7.20 (d, H–C(6)(Dns)); 6.47 (t, H–C(1')); 4.61 (m, H–C(3')); 4.56–4.35 (m, CH₂O); 4.53 (t, CH₂O); 4.22 (m, 2 H–C(5')); 4.16 (m, H–C(4')); 3.88 (br., OH–C(3')); 3.70 (t, SO₂CH₂); 3.14 (t, C–CH₂); 2.88 (t, Me₂N); 2.88–2.76 (t, H–C(2')); 2.60–2.51 (t, H'–C(2')). Anal. calc. for C₃₄H₃₅N₇O₁₁S·1 H₂O (767.8): C 53.19, H 4.86, N 12.77; found: C 52.84, H 4.66, N 12.46.
- **20**: UV (MeOH): 213 (5.03), 259 (4.70), 343 (3.94). 1 H-NMR (CDCl₃): 8.72 (s, H-C(2)); 8.66, 8.60 (2d, 2 H, H-C(2)(Dns)); 8.50 (br., NH); 8.32 (t, 4 H, H-C(4)(Dns), H-C(8)(Dns)); 8.15 (d, 2 H o to NO₂); 8.14 (s, H-C(8)); 7.66–7.54 (m, 4 H, H-C(3)(Dns), H-C(7)(Dns)); 7.42 (d, 2 H m to NO₂); 7.19 (t, 2 H, H-C(6)(Dns)); 6.40 (dd, H-C(1')); 5.16 (d, H-C(3')); 4.56–4.41 (m, 3 CH₂O); 4.29 (m, 2 H-C(5')); 4.15 (m, H-C(4')); 3.71 (m, 2 SO₂CH₂); 3.15 (t, C-CH₂); 2.92–2.80 (m, H-C(2')); 2.87 (s, 2 Me₂N); 2.59–2.51 (dd, H'-C(2')). Anal. calc. for C₄₉H₅₀N₈O₁sS₂·2 H₂O (1091.2): C 53.94, H 4.99, N 10.27; found: C 53.79, H 4.65, N 10.11.
- 9. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy- N^2 -[2-(4-nitrophenyl)ethoxycarbonyl]- O^6 -[2-(4-nitrophenyl)ethyl]guanosine (21) and 3',5'-Bis-O-(2-dansylethoxycarbonyl)-2'-deoxy- N^2 -[2-(4-nitrophenyl)ethoxycarbonyl]- O^6 -[2-(4-nitrophenyl)ethyl]guanosine (22). As described in Exper. 6, with 2 g (3.28 mmol) of 12 [35] [36] and 1.61 g (4.27 mmol) of 8 in 30 ml of dry pyridine. Purification was achieved by FC (silica gel, 18×4 cm, $CH_2Cl_2/MeOH$ 100:0 (0.3 l), 99:1 (1 l; 22), 98:2 (1.5 l; 21), and 97:3 (0.6 l; 21)). After repurification of 22 (CH_2Cl_2 soln.) by FC (silica gel, 9×3 cm, toluene/AcOEt 2:1 (250 ml), toluene/AcOEt 1:1 (100 ml), toluene/AcOEt 1:2 (150 ml; 22), and AcOEt (100 ml; 22)), 2.39 g (80%) of 21 and 0.34 g (8%) of 22 were obtained as yellow amorphous solids.
- **21**: UV (MeOH): 214 (4.91), 262 (4.63), 341 (3.67). 1 H-NMR (CDCl₃): 8.59 (d, H–C(2)(Dns)); 8.30 (t, H–C(4)(Dns), H–C(8)(Dns)); 8.15 (m, 4 H o to NO₂); 7.91 (s, H–C(8)); 7.64–7.55 (m, H–C(3)(Dns), H–C(7)(Dns)); 7.50 (br., NH); 7.49, 7.41 (m, 4 H m to NO₂); 7.20 (d, H–C(6)(Dns)); 6.43 (t, H–C(1')); 4.77 (t, H–C(3'), CH₂O from O^6 -npe); 4.54–4.37 (m, 2 CH₂O); 4.35–4.18 (2dd, 2 H–C(5')); 4.16 (m, H–C(4')); 3.69 (t, SO₂CH₂); 3.62 (br., OH–C(5')); 3.29, 3.12 (2t, 2 C–CH₂); 2.89 (t, Me₂N); 2.89–2.78 (t, H–C(2')); 2.54–2.47 (t,

- H'-C(2')). Anal. calc. for $C_{42}H_{42}N_8O_{14}S\cdot 0.5$ H_2O (923.9): C 54.60, H 4.69, N 12.13; found: C 54.54, H 4.67, N 11.94.
- **22**: UV (MeOH): 214 (5.09), 256 (4.76), 343 (3.90). ¹H-NMR (CDCl₃): 8.65, 8.59 (2*d*, 2 H, H–C(2)(Dns)); 8.34–8.27 (*m*, 4 H, H–C(4)(Dns), H–C(8)(Dns)); 8.15 (*m*, 4 H o to NO₂); 7.89 (*s*, H–C(8)); 7.66–7.53 (*m*, 4 H, H–C(3)(Dns), H–C(7)(Dns)); 7.52 (br., *s*, NH); 7.50, 7.42 (*m*, 4 H m to NO₂); 7.19 (*t*, 2 H, H–C(6)(Dns)); 6.23 (*t*, H–C(1')); 5.27 (*d*, H–C(3')); 4.79 (*t*, CH₂O from O⁶-npe); 4.54–4.41 (*m*, 3 CH₂O); 4.35 (*m*, 2 H–C(5')); 4.14 (*m*, H–C(4')); 3.69 (*m*, 2 SO₂CH₂); 3.30, 3.12 (2*t*, 2 C–CH₂); 3.10–3.00 (*m*, H–C(2')); 2.87 (*s*, 2 Me₂N); 2.49–2.42 (*m*, H′–C(2')). Anal. calc. for C₅₇H₅₇N₉O₁₈S₂ (1220.3): C 56.11, H 4.71, N 10.33; found: C 55.79, H 4.76, N 10.01.
- 10. Bis(2-dansylethyl) Carbonate (23). In Exper. 6-9, 23 was formed as a by-product in variable yields up to 20% and isolated by evaporating the first eluating fractions from FC to a small volume and crystallization by addition of MeOH. Yellow crystals. M.p. 195-196°. UV (CHCl₃): 257 (4.44), 351 (3.88). ¹H-NMR (CDCl₃): 8.59 (d, 2 H, H-C(2)); 8.26 (t, 4 H, H-C(4), H-C(8)); 7.63-7.52 (m, 4 H, H-C(3), H-C(7)); 7.19 (d, 2 H, H-C(6)); 4.27 (t, 2 CH₂O); 3.58 (t, 2 SO₂CH₂); 2.89 (s, 2 Me₂N). Anal. calc. for C₂₉H₃₂N₂O₇S₂ (584.7): C 59.57, H 5.52, N 4.79; found: C 58.96, H 5.55, N 4.84.
- 11. 2-(4-Nitrophenyl)ethyl Chloro (diisopropylamido) phosphite (24) [37]. Under N_2 , 16.7 g (0.1 mol) of 2-(4-nitrophenyl)ethanol were added in small portions at -30° within 1.5 h to 56 ml (0.63 mol) of distilled PCl₃ in abs. Et₂O (150 ml). Thereafter, the mixture was stirred at r.t. for 3 h. Then, volatiles were removed under high vacuum. To the residue, 17.3 g (0.1 mol) of N_i N-diisopropyl(trimethylsilyl)amine were added dropwise at 0° and under N_2 . The mixture was stirred for 30 min at 0° and 20 h at r.t. Thereafter, volatiles were removed under high vacuum: 32.2 g (97%) of 24. Slightly brownish solid which was used in this form for further reactions. 31 P-NMR (CH₂Cl₂): 181.6.
- 12. 2-(4-Nitrophenyl)ethyl Chloro(diethylamido)phosphite (25) [38]. As described in Exper. 11, with 5 g (30 mmol) of 2-(4-nitrophenyl)ethanol and 19.2 ml (0.22 mol) of PCl_3 in abs. THF (100 ml) at -40° . The mixture was stirred at r.t. for 2 h, then evaporated. N,N-Diethyl(trimethylsilyl)amine (4.36 g, 30 mmol) in abs. THF (40 ml) was added dropwise at -20° to the residue in abs. THF (100 ml) and the mixture stirred for 1 h at 35° and then evaporated: 8.20 g (90%) of 25. Slightly brownish oil which was used in this form for further reactions. 31 P-NMR (CDCl₃): 177.3.
- 14. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]cytidine 3'-[2-(4-Nitrophenyl)ethyl N,N-Diisopropylphosphoramidite] (27). As described in Exper. 13, with 363 mg (0.5 mmol) of 16. Purification by FC (silica gel, 9 × 2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 3:1 (100 ml), 2:1 (100 ml), and 1:1 (100 ml), 27) gave 316 mg (62%) of 27. Yellow foam. UV (MeOH): 212 (4.87), 254 (4.54), 285 (sh, 4.37), 342 (3.61). ¹H-NMR (CD₃CN): 8.57 (d, H-C(2)(Dns)); 8.47 (br., NH); 8.23 (t, H-C(4)(Dns), H-C(8)(Dns)); 8.15-8.07 (m, 4 H o to NO₂); 7.76 (2d, H-C(6)); 7.60, 7.59 (2t, H-C(3)(Dns), H-C(7)(Dns)); 7.50, 7.45 (2d, 4 H m to NO₂); 7.21 (2d, H-C(6)(Dns)); 7.00 (m, H-C(5)); 6.05 (t, H-C(1')); 4.40 (2t, 2 CH₂OCO); 4.25 (m, H-C(3')); 4.15-3.76 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.71 (t, SO₂CH₂); 3.51 (m, 2 Me₂CH); 3.08, 2.98 (2t, C-CH₂); 2.82 (s, Me₂N); 2.50-2.38 (m, H-C(2')); 2.01-1.88 (m, H'-C(2')); 1.13-1.06 (m, 2 Me₂CH). ³¹P-NMR (CD₃CN): 147.91, 147.77. Anal. calc. for C₄₇H₅₆N₇O₁₅PS (1022.1): C 55.23, H 5.52, N 9.59; found: C 54.81, H 5.50, N 9.67.
- 15. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N⁶-[2-(4-nitrophenyl)ethoxycarbonyl]adenosine 3'-[2-(4-Nitrophenyl)ethyl N, N-Diisopropylphosphoramidite] (28). As described in Exper. 13, with 375 mg (0.5 mmol) of 19. Purification by FC (silica gel, 9×2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 3:1

- (100 ml), 2:1 (100 ml), and 1:1 (100 ml, **28**)) gave 345 mg (66%) of **28**. Yellow foam. UV (MeOH): 206 (4.95), 265 (4.67), 341 (3.65). 1 H-NMR (CD₃CN): 8.91 (br., NH); 8.57, 8.55 (2s, H-C(2)); 8.52 (2d, H-C(2)(Dns)); 8.23-8.02 (m, H-C(8)(Dns), H-C(8)(Dns), 4 H o to NO₂); 7.60-7.43 (m, H-C(3)(Dns), H-C(7)(Dns), 4 H m to NO₂); 7.19 (2d, H-C(6)(Dns)); 6.32 (m, H-C(1')); 4.59 (m, H-C(3')); 4.45, 4.44, 4.32 (3 t, 2 CH₂OCO); 4.20-3.78 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.67 (t, SO₂CH₂); 3.62-3.48 (m, Me₂CH); 3.10, 3.00, 2.99 (3 t, 2 C-CH₂); 2.79 (2s, Me₂N); 2.79-2.63 (m, H-C(2')); 2.52-2.41 (m, H'-C(2')); 1.16-1.09 (m, 2 m₂CH). 31 P-NMR (CD₃CN): 147.77, 147.69. Anal. calc. for C₄₈H₅₆N₉O₁₄PS (1046.1): C 55.11, H 5.40, N 12.05; found: C 54.88, H 5.45, N 12.02.
- 16. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N²-[2-(4-nitrophenyl)ethoxycarbonyl]-O³-[2-(4-nitrophenyl)ethyl] guanosine 3'-[2-(4-Nitrophenyl)ethyl] N,N-Diisopropylphosphoramidite] (29). As described in Exper. 13, with 457 mg (0.5 mmol) of 21. Purification by FC (silica gel, 9×2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 4:1 (100 ml) and 3:1 (100 ml, 29)) gave 272 mg (45%) of 29. Yellow foam. UV (MeOH): 213 (4.93), 264 (4.69), 339 (3.68). ¹H-NMR (CD₃CN): 8.51 (d, H-C(2)(Dns)); 8.31 (s, NH); 8.24-7.97 (m, H-C(4)(Dns), H-C(8)(Dns), 6 H o to NO₂); 7.88 (s, H-C(8)); 7.61-7.39 (m, H-C(3)(Dns), H-C(7)(Dns), 6 H m to NO₂); 7.19 (2d, H-C(6)(Dns)); 6.19 (m, H-C(1')); 4.77 (m, CH₂O from O³-npe); 4.70 (m, H-C(3')); 4.41-4.30 (m, 2 CH₂OCO); 4.26-3.78 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.66 (2t, SO₂CH₂); 3.53 (m, 2 Me₂CH); 3.27, 3.10, 2.97 (3 m, 3 C-CH₂); 2.90-2.71 (m, H-C(2')); 2.79 (s, Me₂N); 2.42-2.30 (m, H'-C(2')); 1.18-1.07 (m, 2 Me₂CH). ³P-NMR (CD₃CN): 147.86. Anal. calc. for $C_{56}H_{63}N_{10}O_{17}PS$ (1211.2): C 55.53, H 5.24, N 11.56; found: C 55.23, H 5.35, N 11.41.
- 17. 5'-O-(2-Dansylethoxycarbonyl) thymidine 3'-[2-(4-Nitrophenyl) ethyl N, N-Diethylphosphoramidite] (30). As described in Exper. 13, with 274 mg (0.5 mmol) of 13, 0.52 ml (3 mmol) of abs. (i-Pr)₂EtN, 0.31 g (1 mmol) of 25, and 4 ml of dry THF (1.5 h at r.t.). Purification by FC (silica gel, 6 × 2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 4:1 (60 ml), 3:1 (60 ml), and 2:1 (60 ml, 30)) gave 277 mg (68%) of 30. Yellow foam. UV (MeOH): 213 (4.74), 260 (4.47), 343 (3.61). ¹H-NMR (CD₃CN): 9.01 (br., s, NH); 8.58 (2d, H-C(2)(Dns)); 8.27-8.19 (m, H-C(4)(Dns), H-C(8)(Dns)); 8.11 (d, 2 H o to NO₂); 7.65-7.57 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.46 (d, 2 H m to NO₂); 7.24 (d, H-C(6)(Dns)); 7.11 (t, H-C(6)); 6.10 (t, H-C(1')); 4.40 (m, CH₂OCO); 4.28 (m, H-C(3')); 4.22-3.79 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.72 (t, SO₂CH₂); 3.09-2.88 (m, C-CH₂, 2 MeCH₂); 2.83 (2s, Me₂N); 2.23-2.12 (m, H-C(2')); 1.99-1.86 (m, H'-C(2')); 1.72, 1.70 (2d, Me); 1.02-0.94 (m, 2 MeCH₂). ³¹P-NMR (CD₃CN): 148.31, 147.86. Anal. calc. for C₃₇H₄₆N₃O₁₂PS (815.8): C 54.47, H 5.68. N 8.58; found: C 54.29, H 5.74, N 8.58.
- 18. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]cytidine 3'-[2-(4-Nitrophenyl)ethyl N, N-Diethylphosphoramidite] (31). As described in Exper. 17, with 363 mg (0.5 mmol) of 16. Purification by FC (silica gel, 5 × 2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 3:1 (50 ml), 2:1 (50 ml), and 1:1 (50 ml, 31)) gave 295 mg (59%) of 31. Yellow foam. UV (MeOH): 212 (4.85), 255 (4.54), 284 (sh, 4.38), 340 (3.64). ¹H-NMR (CD₃CN): 8.56 (d, H-C(2)(Dns)); 8.41 (br., NH); 8.25-8.14 (m, H-C(4)(Dns), H-C(8)(Dns)); 8.15-8.08 (m, 4 H o to NO₂); 7.75, 7.74 (2d, H-C(6)); 7.62-7.55 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.52-7.43 (m, 4 H m to NO₂); 7.21 (d, H-C(6)(Dns)); 6.99 (br., d, H-C(5)); 6.03 (m, H-C(1')); 4.40 (t, 2 CH₂OCO); 4.25 (m, H-C(3')); 4.15-3.75 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.71 (t, SO₂CH₂); 3.08 (t, C-CH₂); 3.04-2.89 (m, C-CH₂); 2.82 (s, Me₂N); 2.48-2.33 (m, H-C(2')); 2.02-1.87 (m, H'-C(2')); 1.02-0.93 (m, 2 MeCH₂). ³¹P-NMR (CD₃CN): 148.25, 147.83. Anal. calc. for C₄₅H₅₂N₇O₁₅PS (994.0): C 54.38, H 5.27, N 9.86; found: C 54.08, H 5.39, N 9.86.
- 19. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N 6 -[2-(4-nitrophenyl)ethoxycarbonyl]adenosine 3'-[2-(4-Nitrophenyl)ethyl N, N-Diethylphosphoramidite] (32). As described in Exper. 17, with 375 mg (0.5 mmol) of 19. Purification by FC (silica gel, 5×2 cm, CH₂Cl₂/petroleum ether soln., then quickly petroleum ether/acetone 3:1 (50 ml), 2:1 (50 ml), and 1:1 (100 ml, 32)) gave 325 mg (64%) of 32. Yellow foam. UV (MeOH): 210 (4.89), 264 (4.66), 343 (3.65). 1 H-NMR (CD₃CN): 8.92 (br., s, NH); 8.56, 8.55 (2s, H-C(2)); 8.51 (d, H-C(2)(Dns)); 8.22-8.15 (m, H-C(4)(Dns), H-C(8)(Dns)); 8.11-8.04 (m, H-C(8), 4 H o to NO₂); 7.59-7.50 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.48-7.42 (m, 4 H m to NO₂); 7.18 (d, H-C(6)(Dns)); 6.30 (t, H-C(1')); 4.59 (m, H-C(3')); 4.43 (2t, CH₂OCO); 4.31 (2t, CH₂OCO); 4.34-3.72 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.66 (t, SO₂CH₂); 3.08 (t, C-CH₂); 3.02-2.93 (m, C-CH₂, 2 MeCH₂); 2.78 (2s, Me₂N); 2.77-2.64 (m, H-C(2')); 2.51-2.38 (m, H'-C(2')); 0.99 (t, 2 Me). 13 P-NMR (CD₃CN): 147.98, 147.77. Anal. calc. for $C_{46}H_{52}N_{9}O_{14}$ PS (1018.0): C 54.27, H 5.15, N 12.38; found: C 53.82, H 5.22, N 11.99.
- 20. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy- N^2 -[2-(4-nitrophenyl)ethoxycarbonyl]- O^6 -[2-(4-nitrophenyl)ethyl]guanosine 3'-[2-(4-Nitrophenyl)ethyl N,N-Diethylphosphoramidite] (33). As described in Exper. 17, with 457 mg (0.5 mmol) of 21. Purification by FC (silica gel, 5×2 cm, CH₂Cl₂/petroleum ether soln., then quickly

- petroleum ether/acetone 4:1 (50 ml), 3:1 (50 ml), 2:1 (50 ml; 33), and 1:1 (50 ml; 33)) gave 340 mg (57%) of 33. Yellow foam. UV (MeOH): 214 (4.93), 264 (4.70), 340 (3.66). 1 H-NMR (CD₃CN): 8.50 (d, H-C(2)(Dns)); 8.31 (s, NH); 8.23-7.99 (m, H-C(4)(Dns), H-C(8)(Dns), 6 H o to NO₂); 7.87 (s, H-C(8)); 7.60-7.38 (m, H-C(3)(Dns), H-C(7)(Dns), 6 H m to NO₂); 7.18 (d, H-C(6)(Dns)); 6.17 (m, H-C(1')); 4.75 (m, CH₂O from O^6 -npe, H-C(3')); 4.40-4.28 (m, 2 CH₂OCO); 4.25-3.78 (m, CH₂OP, H-C(4'), 2 H-C(5')); 3.66 (t, SO₂CH₂); 3.27/3.26 (2t, C-CH₂ from O^6 -npe); 3.08 (t, C-CH₂); 3.03-2.89 (t, C-CH₂, MeCH₂); 2.86-2.72 (t, H-C(2')); 2.78 (t, Me₂N); 2.39-2.31 (t, H'-C(2')); 1.02-0.93 (t, MeCH₂). t³¹P-NMR (CD₃CN): 148.03, 147.97. Anal calc. for C₅₄H₅₉N₁₀O₁₇PS (1183.2): C 54.82, H 5.03, N 11.84; found: C 54.25, H 5.12, N 11.50.
- 21. 5'-O-(2-Dansylethoxycarbonyl)-3'-O-succinylthymidine (34). In 5 ml of dry CH₂Cl₂, 548 mg (1 mmol) of 13, 0.2 g (2 mmol) of succinic anhydride, and 0.16 g (1.3 mmol) of 4-(dimethylamino)pyridine were stirred at r.t. for 2 h. Then, the mixture was diluted with CH₂Cl₂ (50 ml) and washed with sat. NaHCO₃ soln. (50 ml). To obtain a complete phase separation, 10% citric acid was added. The aq. phase was extracted with CH₂Cl₂ (2 × 25 ml), the combined org. layer washed with 10% citric acid (50 ml), the aq. phase extracted with CH₂Cl₂ (25 ml), and then the combined org. layer dried (Na₂SO₄) and evaporated: 0.61 g (94%) of 34. Yellow amorphous solid. UV (MeOH): 214 (4.71), 257 (4.36), 346 (3.63). ¹H-NMR ((D₆)DMSO): 12.29 (br., COOH); 11.38 (s, NH); 8.53 (d, H-C(2)(Dns)); 8.18 (t, H-C(4)(Dns), H-C(8)(Dns)); 7.66 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.34 (s, H-C(6)); 7.26 (d, H-C(6)(Dns)); 6.14 (t, H-C(1')); 5.09 (m, H-C(3')); 4.38 (m, CH₂O); 4.15 (m, 2 H-C(5')); 4.02 (m, H-C(4')); 3.87 (t, SO₂CH₂); 2.81 (s, Me₂N); 2.52 (m, CH₂CH₂); 2.23 (t, 2 H-C(2')); 1.68 (s, Me). Anal. calc. for C₂₉H₃N₃O₁₂S (647.7): C 53.78, H 5.14, N 6.49; found: C 53.78, H 5.34, N 6.11.
- 22. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N⁴-[2-(4-nitrophenyl)ethoxycarbonyl]-3'-O-succinylcytidine (35). As described in Exper. 21, with 363 mg (0.5 mmol) of 16. Workup yielded 384 mg (93%) of 35 as a yellow amorphous solid which was used in this form for reaction with the support. An anal. sample was obtained by FC (silica gel, 6×2 cm, CH₂Cl₂/MeOH 100:0 (50 ml), 99:1 (50 ml), 98:2 (50 ml), and 97:3 (50 ml)). UV (MeOH): 214 (4.79), 247 (4.47), 286 (sh, 4.18), 343 (3.61). 1 H-NMR ((D₆)DMSO): 12.20 (br., COOH); 10.90 (br., NH); 8.52 (d, H-C(2)(Dns)); 8.18-8.12 (m, H-C(4)(Dns), H-C(8)(Dns)); 8.15 (d, 2 H o to NO₂); 7.90 (d, H-C(6)); 7.69-7.57 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.59 (d, 2 H o to NO₂); 7.23 (d, H-C(6)(Dns)); 6.98 (d, H-C(5)); 6.09 (d, H-C(1')); 5.10 (d, H-C(3')); 4.40-4.33 (d, 2 CH₂O); 4.16 (d, H-C(4'), 2 H-C(5')); 3.86 (d, SO₂CH₂); 3.07 (d, C-CH₂); 2.79 (d, Me₂N); 2.58-2.38 (d, CH₂CH₂, H-C(2')); 2.23-2.11 (d, H'-C(2')). Anal. calc. for C₃₇H₃₉N₅O₁₅S·0.5 H₂O (834.8): C 53.23, H 4.83, N 8.39; found: C 53.25, H 4.79, N 8.49.
- 23. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N 6 -[2-(4-nitrophenyl)ethoxycarbonyl]-3'-O-succinyladenosine (36). As described in Exper. 21, with 375 mg (0.5 mmol) of 19. On washing with sat. NaHCO $_3$ soln., no 10% citric acid was needed to achieve complete phase separation. Further workup yielded 404 mg (95%) of 36. Purification for anal. characterization as described in Exper. 22. UV (MeOH): 211 (4.83), 263 (4.57), 342 (3.64). 1 H-NMR ((D_6) DMSO): 12.32 (br., COOH); 10.63 (br. s, NH); 8.62, 8.55 (2s, H-C(2), H-C(8)); 8.48 (d, H-C(2)(Dns)); 8.17 (t, H-C(4)(Dns), H-C(8)(Dns)); 8.16 (d, 2 H o to NO $_2$); 7.66-7.58 (m, H-C(3)(Dns), H-C(7)(Dns)); 7.09 (d, 2 H m to NO $_2$); 7.23 (d, H-C(6)(Dns)); 6.44 (t, H-C(1')); 5.35 (m, H-C(3')); 4.38, 4.32 (2t, 2 CH $_2$ O); 4.24-4.13 (m, H-C(4'), 2 H-C(5')); 3.84 (t, SO $_2$ CH $_2$); 3.10 (t, C-CH $_2$); 3.10-3.03 (t, H-C(2')); 2.78 (t, Me $_2$ N); 2.56 (t, CH $_2$ CH $_2$, H'-C(2')). Anal. calc. for C $_3$ 8H $_3$ 9N $_7$ O $_1$ 4S·0.75 H $_2$ O (863.4): C 52.86, H 4.73, N 11.36; found: C 52.87, H 4.71, N 11.37.
- 24. 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-N²-[2-(4-nitrophenyl)ethoxycarbonyl]-O²-[2-(4-nitrophenyl)ethyl]-3'-O-succinylguanosine (37). As described in Exper. 21, with 458 mg (0.5 mmol) of 21. On washing with sat. NaHCO₃ soln., no 10% citric acid was needed to achieve complete phase separation. Further workup yielded 450 mg (89%) of 37. Purification for anal. characterization as described in Exper. 22. UV (MeOH): 214 (4.89), 262 (4.62), 343 (3.66). ¹H-NMR ((D₆)DMSO): 12.31 (br., COOH); 10.38 (s, NH); 8.47 (d, H-C(2)(Dns)); 8.29 (s, H-C(8)); 8.19-8.12 (m, H-C(4)(Dns), H-C(8)(Dns)); 8.15 (d, 4 H o to NO₂); 7.61 (t, H-C(3)(Dns), H-C(7)(Dns), 4 H m to NO₂); 7.22 (d, H-C(6)(Dns)); 6.32 (t, H-C(1')); 5.38 (m, H-C(3')); 4.75 (t, CH₂O from O⁶-npe); 4.39-4.26 (m, 2 CH₃O); 4.23-4.14 (m, H-C(4'), 2 H-C(5')); 3.82 (t, SO₂CH₂); 3.28 (t, C-CH₂ from O⁶-npe); 3.19-3.04 (m, H-C(2')); 3.09 (t, C-CH₂); 2.77 (s, Me₂N); 2.58-2.43 (m, CH₂CH₂, H'-C(2')). Anal. calc. for C₄₆H₄₆N₈O₁₇S (1015.0): C 54.44, H 4.57, N 11.04; found: C 54.07, H 4.63, N 10.98.
- 25. (Long-chain-alkyl) methylamine Controlled-Pore Glass (LCAMA-CPG; 38). A mixture of 5 g of glyceryl-CPG 500 Å (120-200 mesh, Fluka) and 5 g (26.3 mmol) of N-(phenoxycarbonyl)-1H-tetrazolide [47] [48] was shaken in dry CH_2Cl_2 (70 ml) at r.t. for 20 h. Then, the material was collected in a glass suction filter and washed with CH_2Cl_2 (4 ×) by taking up the CPG in CH_2Cl_2 (50 ml) and suction-filtering. Then, the CPG was reacted with 4.4 ml (25.4 mmol) of N,N'-dimethylhexane-1,6-diamine in dry CH_2Cl_2 (50 ml) at 60° for 2 h. The LCAMA-CPG (38) was isolated in a glass suction filter and washed with MeOH, DMF, MeOH, acetone, and Et_2O .

- 26. Derivatives 39-42 of LCAMA-CPG 500 Å (38) and of 5'-O-(2-Dansylethoxycarbonyl)-2'-deoxy-3' O-succinylnucleosides 34-37. To a mixture of 200 mg of 38, 45 μ mol of 34 (29 mg), 35 (37 mg), 36 (38 mg), or 37 (46 mg), and 30 mg (91.4 μ mol) of TOTU, 3 ml of abs. MeCN and 19 μ l (180 μ mol) of N-methylmorpholine were added. After a short ultrasonic treatment (30 s) and 2 h standing in the dark at r.t., the CPG material was collected in a glass suction filter and washed with MeOH, DMF, MeOH, and Et₂O. Capping-procedure: The nucleoside-functionalized CPG, 50 mg (0.41 mmol) of 4-(dimethylamino)pyridine, 10 ml of abs. pyridine, and 1 ml (10.6 mmol) of Ac₂O were kept in the dark for 45 min. at r.t. Thereafter, 39-42, resp., were collected in a glass suction filter and washed with MeOH, DMF, MeOH, and Et₂O. Determination of loading: A defined amount of 39, 40, 41, or 42 (5-10 mg) was weighed in a 1-ml microcuvette (d=1 cm). Then, 0.1m DBU in MeCN (500 μ l) was added. After 1 min, the soln. was neutralized with 0.1m AcOH in MeCN (500 μ l). Thereafter, the absorbance at 345 nm was measured against 0.05m DBU-acetate in MeCN. By considering log $\varepsilon=3.63$ of the formed 5-(dimethylamino)naphthalen-1-yl vinyl sulfone, the loading L [μ mol/g] can be calculated by the formula $L=234.75 \cdot A/m$ (A= absorbance at 345 nm; m= weighed CPG (39-42) in mg): L (39) = 19, L (40) = 19, L (41) = 21, and L (42) = 21 μ mol/g.
- 27. Assembly of Oligodeoxynucleotides. Syntheses were carried out using an Applied Biosystems 380B DNA synthesizer. Nucleoside-functionalized CPG material (39–42; 0.6 μmol) was packed into a small ABI column, and cycles of nucleotide addition were carried out by a programmed series of reagent and solvent washes based on recommended procedures with the following main steps: 1) 5'-O-Dnseoc Deprotection: 0.1 m DBU in MeCN delivered in 2 30-s and 8 10-s bursts with intermediate 1-s reverse flushes. The eluate from this step was collected and the absorbance at 350 nm measured to determine the condensation yields. 2) Coupling: 0.1 m phosphoramidite (P) and 0.5 m 1H-tetrazole (T) in dry MeCN delivered in 5 alternating bursts (8 s T, 4 s P + T, 3 s T, 3 s P + T, 3 s T) with a subsequent wait time of 40 s. 3) Capping: Ac₂O/2,6-dimethylpyridine/THF 1:1:8 and 1-methyl-1H-imidazole/THF 16:84 delivered in one 20-s burst with a subsequent wait time of 30 s. 4) Oxidation: 0.05 m I₂ in THF/H₂O/pyridine 7:2:1 delivered in one 23-s burst with a subsequent wait time of 30 s.

Then a cleavage programme was carried out: 1) npe/npeoc Deprotection: 0.5M or 1M DBU in MeCN delivered in one 180-s burst with a consecutive wait time of 3600 s and 5 times one 120-s burst followed by another wait time of 5400 s (total wait time 10 h). 2) Cleavage from the support: conc. NH₃ delivered in one 18-s burst with a consecutive wait time of 2400 S (5 times, total wait time 3 h 20 min). The ammoniacal oligonucleotide containing soln. was collected and, after determination of the isolated amount of oligonucleotide by measurement of the absorbance at 260 nm, was lyophilized in a *Speed-vac* concentrator under high vacuum.

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